

INTRODUCTION

The DNA Amplification Kit is designed to contain high quality Taq DNA Polymerases, buffers, dNTPs mix, and nuclease-free water as a complete set for user's convenience in performing DNA amplification experiments. The kit allows for up to 200 applications or more in a 50 μ l PCR reaction. DNA and primers are also provided as a positive control for users to carry out PCR using the recommended parameters as shown in this manual.

The kit is also supplied with DNA ladders (ready-to-use) for up to 100 applications to determine the size of PCR products or other double-stranded DNA fragments during gel electrophoresis.

KIT COMPONENTS

PCR Amplification Reagents

| Taq DNA Polymerase (5u/μl) | 500u |
|--|---------|
| 10X ViBuffer A | 2ml |
| (500mM KCl, 100mM Tris-HCl (pH 9.1) and 0.1% Triton X-100) | |
| 10X ViBuffer S | 1ml |
| (160Mm (NH ₄) ₂ SO ₄ , 500mM Tris-HCl (pH9.2), 17.5mM MgCl ₂ and 0.1% Triton X-100) | |
| 50mM MgCl ₂ | 1ml |
| 2mM dNTPs Mix | 1ml |
| Control DNA (5ng/µl) | 100ng |
| 10μM Forward primer | 25µ1 |
| 10μM Reverse primer | 25µ1 |
| Nuclease-free water | 1ml X 2 |

DNA Ladder (ready-to-use)

 $\begin{array}{ccc} VC \ 100 bp \ plus \ (0.1 \mu g/\mu l) & 100 \ applications \\ VC \ 1kb \ (0.1 \mu g/\mu l) & 100 \ applications \\ 6X \ loading \ dye & 1ml \end{array}$

STORAGE & STABILITY

Store all components at -20°C

Kit components are guaranteed to be stable for 2 year from the date of manufacture.

PCR PROTOCOL (Control DNA)

- 1. Gently mix all solutions after thawing. Keep solutions on ice from this point onwards.
- 2. Add the following reagents into a PCR tube, on ice.

| Reagent | Quantity (µl) | Final Concentration |
|----------------------------|---------------|----------------------------|
| Water, nuclease-free | 38.1 | - |
| 10X ViBuffer A | 5.0 | 1X |
| 2mM dNTPs mix | 2.0 | 0.08mM |
| 50mM MgCl ₂ | 1.5 | 1.5mM |
| 10uM Forward Primer | 1.0 | 0.2μΜ |
| 10uM Reverse Primer | 1.0 | 0.2μΜ |
| Control DNA (5ng/µl) | 1.0 | 5ng |
| Taq DNA Polymerase (5u/µl) | 0.4 | 2unit |
| Total Volume | 50.0 | |

- 3. Gently mix the PCR reagents. Briefly centrifuge the tubes to collect the contents at the bottom of the tube.
- 4. Perform DNA amplification using the following program:

| Segment | No. of cycles | Temperature | Duration |
|---------|---------------|-------------|----------|
| 1 | 1 | 95°C | 3 min |
| 2 | 30 | 95°C | 30 sec |
| | | 52°C | 30 sec |
| | | | |
| | | 72°C | 30 sec |
| 3 | 1 | 72°C | 5 min |
| | 1 | 4°C | pause |

5. Run 5μ l of the PCR products along with $0.3-0.5\mu g$ of VC100bp plus DNA ladder in a 1.0% agarose gel. Stain gel with EtBr to visualize DNA bands under UV.

Note: The PCR product of the positive control should provide a 1.4kb DNA fragment size.

TROUBLESHOOTING

PCR Reactions

| Problems | Possibility | Suggestions |
|--|--|--|
| Low yield or no PCR product | Missing component in reaction | Check the reaction components and repeat the reaction. |
| Multiple, non-specific amplification products | Cross contamination of DNA Excessive amounts of enzyme used Excessive amounts of DNA template used Excessive number of cycles Excessive amount of MgCl ₂ used | Use a separate workplace and pipettes for PCR. Wear gloves at all times. Decrease amount of <i>Taq</i> DNA Polymerase in the reaction tube. Decrease amount of DNA template in the reaction tube. Reduce number of cycles. Decrease the concentration of MgCl ₂ in the reaction tube. |
| | Long extension time Pipetting error | Reduce extension time. Perform PCR in reaction master mixes. |
| Smearing of PCR product when viewed after gel electrophoresis | Agarose gel used was not fresh Insufficient amount of MgCl ₂ used Excessive amounts of enzyme used | Repeat electrophoresis with fresh agarose gel. Increase the concentration of MgCl ₂ in the reaction tube. Decrease amount of <i>Taq</i> DNA Polymerase in the reaction tube. |